

# Influenza B Virus: Nucleoprotein (NP) Antibody Inhibition Test

## Product Insert

Cat. No. IBV307-2 (2 x 96 wells)

**For Research Use Only**

### Introduction

Influenza viruses can be divided into three classes, A, B, and C, largely based upon conserved antigenic differences in the internal nucleoprotein. Influenza A virus, typically encountered more frequently than types B and C, and associated with the majority of serious epidemics, can be further subdivided into strains or subtypes based on antigenic differences in the external hemagglutinin proteins (H1-H18) and neuraminidase proteins (N1-N11).

Historically, human influenza A virus infections have been associated with H1N1, N2N2, and H3N2 subtypes of influenza A, although a 1997 outbreak in Hong Kong was identified as an H5N1 subtype. This outbreak was not only significant because it resulted in multiple human infections and deaths, but it also represented the first known demonstration of avian influenza virus transmission to humans.

While influenza A virus will infect a wide variety of species, influenza B virus is predominantly a human pathogen, although it has been found to infect seals. The limited host range of influenza B and a slower rate of mutation than influenza A appears to preclude development of influenza B pandemics, but influenza B is a significant human pathogen and on an individual basis, infection may result in death.

Virusys has developed a highly sensitive and specific enzyme-linked immunosorbent assay (ELISA) for the detection of influenza B nucleoprotein-specific antibodies in serum which may be used for the detection of influenza B NP antibodies in human serum as well as experimental animals.

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The assay utilizes "inhibition of binding" technology and therefore does not require species-specific conjugates. The assay can be completed in less than 1.5 hr., contains only one wash step, and incorporates proprietary diluents that are designed to prevent the development of nonspecific signal derived from sample matrix and/or the nonspecific adsorption of reactive test components. The result is an assay that is both highly sensitive and specific, and capable of being used for experimental evaluations in species other than humans.

### Components

1. IBV Antigen Capture Plate (96 tests) - 2 ea.
2. IBV Antigen Diluent (1x) - 24 ml
3. IBV Antigen Concentrate (1x) - 4 ml
4. IBV Positive Serum Control (1x) - 0.1 ml
5. IBV Negative Serum Control (1x) - 0.1 ml
6. IBV Detection Antibody, HRP-labeled (1x) - 12 ml
7. Wash Buffer (20x)-50 ml
8. Chromagen Solution (1x) - 22 ml
9. Stop Solution (1x) - 22 ml
10. Sample Dilution Tray - 2 ea.

### Storage

Store all kit components at 2-8° C. Crystal formation may occur in the wash buffer concentrate during prolonged storage at 2-8° C. The crystals can be re-dissolved by swirling the bottle in warm tap water.

### Procedure

1. Remove the kit components from storage and allow them to warm to room temperature.
2. Determine the number of test wells needed. Use one well for each sample. In addition, include two wells each for the **Antigen Diluent Control**, **Positive Serum Control** and the **Negative Serum Control**.
3. Based upon the number of wells required for the assay, prepare **IBV Sample Diluent** from the **IBV Antigen Concentrate** and **IBV Antigen Diluent** using a ratio of 1 ml of **Antigen Concentrate** to 5 ml **Antigen Diluent**. For example, if using the entire plate combine 2 ml of **Antigen Concentrate** with 10 ml of **Antigen Diluent**.
4. To begin the assay, transfer 5 µl of each serum control (two wells each) and each sample (one well each) to the appropriate wells of the dilution tray.
5. Add 100 µl of **IBV Sample Diluent** to each sample, positive control, and negative control well. Mix by pipetting up and down several times.
6. Add 100 µl of the **Antigen Diluent** to the appropriate wells (NOTE: Wells containing **Antigen Diluent** DO NOT receive Sample Diluent).
7. Incubate for 10 min. at room temperature.

8. Transfer 50 µl of sample or control to the appropriate wells of the **IBV Antigen Capture Plate**.
9. Cover the plate and incubate for 30 min. at room temperature on a plate shaker set at moderate speed.
10. Add 50 µl of **1x IBV Detection Antibody** to each well. **DO NOT WASH THE PLATE AT THIS TIME.** Cover the plate and incubate for 30 min. on a plate shaker using the same settings (Step 6).
11. Wash the wells 6x with **1x Wash Buffer**.
12. Add 100 µl of **Chromagen** to each well and incubate for 10 min. on a plate shaker.
13. Stop the reaction by the addition of 100 µl of **Stop Solution**.
14. Shake the plate for 10-15 sec. to ensure that the reaction is uniformly stopped and then read the plate in a plate reader using a 450 nm filter.

### Quality Control

1. The negative serum control absorbance values should be  $\geq 0.600$
2. The positive serum control absorbance values should be  $\leq 0.300$ .
3. The diluent control absorbance values should be  $\leq 0.300$ .
4. The mean absorbance value for the positive control serum should be  $\pm 15\%$  of the mean absorbance value for the diluent control.

### Determination of NP Reduction Index and Interpretation of Results

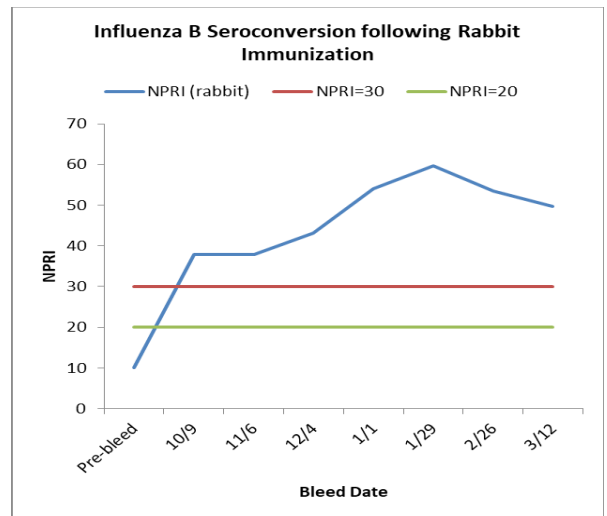
A positive result with the Virusys Influenza B Nucleoprotein Antibody Inhibition Test is dependent upon the presence of NP-specific antibody in the sample. Binding of the antibody in the sample to NP antigen in the sample diluent will result in the complete or partial inhibition of NP binding to the antibody coated on the IBV Antigen Capture Plate. The resulting decrease in NP binding can then be used to calculate a **NP Reduction Index** for each sample.

1. To calculate the NPRI for each sample, it is first necessary to calculate the mean absorbance values for the diluent control and the positive and negative controls.
2. Subtract the mean value of the diluent control from the mean of the positive serum control (maximal NP removal) and from the mean of the negative serum control (minimal NP removal) and the absorbance values for all samples.
3. Calculate the NPRI using the following formula:  
**NPRI = (1-[Delta Sample Abs/Delta NC Abs]) x 100**
4. A NPRI  $>30$  is indicative of the presence of anti-influenza B NP antibody.

5. A NPRI  $<20$  is considered negative for the presence of anti-influenza B NP antibody.
6. A NPRI  $\geq 20$  but  $\leq 30$  represents an equivocal result, suggestive of the presence of NP antibody. Another sample should be obtained in 7-14 days and tested (preferably with the original equivocal sample).

### Performance

Influenza B Seroconversion in Rabbits: Following the acquisition of a pre-bleed sample, a rabbit was immunized with gradient-purified,  $\beta$ -propiolactone-inactivated influenza B virus (Influenza B/Lee/40). Blood samples were collected on the dates indicated in the chart below and tested in the Virusys Influenza B Nucleoprotein Antibody Inhibition Assay. The NP Reduction Index values are shown below. Seroconversion was observed by the first bleed after immunization and the titer rose to an NPRI of 60, e.g., 60% reduction in NP binding, and then declined slightly.



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